



Determination of Fatty Acid and Essential Oil Constituents and Biological Activities on *Ranunculus pedatus* Subsp. *pedatus*

T. ERDOGAN^{1,*}, T. GONENC¹, B. DEMIRCI², B. KIVÇAK¹ and K.H.C. BASER^{2,3,4}

¹Department of Pharmacognosy, Faculty of Pharmacy, Ege University, Bornova, Izmir, Turkey

²Department of Pharmacognosy, Faculty of Pharmacy, Anadolu University, Eskisehir, Turkey

³Botany and Microbiology Department, College of Science, King Saud University, Riyadh, Saudi Arabia

⁴Technology Transfer Office, Bahcesehir University, Istanbul, Turkey

*Corresponding author: Fax: +90 232 3885258; Tel: +90 232 3113965; E-mail: tugce.fafal@ege.edu.tr; tugceerfa@hotmail.com

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In this study, the fatty-acid profile, essential-oil composition and cytotoxic and antioxidant activities of the aerial parts of *Ranunculus pedatus* Waldst. & Kit. subsp. *pedatus* have been examined. Linoleic acid (24.47 %) and palmitoleic acid (18.94 %) were identified as major components by GC-FID and GC/MS analysis. Unsaturated fatty acids were found in higher amounts than saturated fatty acids. The essential oil composition was also analyzed by GC and GC/MS and 29 components were characterized. Protoanemonin (14.3 %), phytol (13.7 %), α -bisabolol (7.9 %), (Z)-3-hexenol (7.8 %) and octanol (6.7 %) were determined as the main constituents of the essential oil. Ethanol and chloroform extracts were investigated for the cytotoxic activity against PC3, HeLa and Vero human cancer cell lines and both of the extracts were found to be effective on HeLa. The IC₅₀ values were determined as 47 and 78.4 mg/mL for methanol and chloroform extracts respectively. Antioxidant activity of chloroform and ethanol extracts were evaluated by DPPH assay. The total phenolics and flavonoid contents of the extracts were determined by Folin-Ciocalteu and aluminium chloride methods, respectively. The ethanol extract was found to possess 67.54 % DPPH inhibition. The results obtained in the antioxidant activity assay were in positive correlation with the total phenolics and flavonoid contents of the extracts.

Keywords: *Ranunculus pedatus* subsp. *pedatus*, Fatty acid, Essential oil, Cytotoxic activity, Antioxidant activity.

INTRODUCTION

Ranunculus genus with the greatest diversity within the family Ranunculaceae is represented by 600 species¹. There are 84 wild-growing species belonging to this genus in Turkey¹⁻³. *Ranunculus* species have been used in traditional medicine for wound healing⁴, antihemorrhoidal⁵⁻⁷, maturation of abscess^{8,9}, against rheumatism⁸ and for the treatment of tuberculosis¹⁰.

This genus have previously been investigated for its flavonoids¹¹⁻¹⁵, saponins^{16,17}, alkaloids^{18,19}, fatty and organic acids²⁰⁻²². In previous studies, *Ranunculus* species have been shown to possess antimicrobial²³⁻²⁵, cytotoxic¹⁵, anti-inflammatory^{14,26-28}, antioxidant^{24,29,30} and antiprotozoal³¹.

The aim of the present study is to evaluate the fatty acid, essential oil composition, cytotoxic and antioxidant activities of *Ranunculus pedatus* subsp. *pedatus* growing wild in Manisa, Turkey. The essential oil composition of *R. arvensis* and *R. constantinopolitanus*³² and antioxidant activities of the extracts of *R. marginatus* var. *trachycarpus* and *R. sprunerianus* previously reported²⁴. To the best of our knowledge, no previous work has been reported on the essential oil and fatty acid composition and biological activities of *R. pedatus* subsp. *pedatus*.

EXPERIMENTAL

R. pedatus Waldst. & Kit. subsp. *pedatus* (Ranunculaceae) was collected from Manisa on May 2005. The plant was identified by M. Ali Önür from Ege University, Faculty of Pharmacy, Department of Pharmacognosy and a voucher specimen (No. 1364) is deposited in Herbarium of the Ege University, Faculty of Pharmacy.

Fatty acid analysis: The dried and powdered aerial parts of the plant material (40 g) have been extracted by petroleum ether (250 mL) for 6 h at 60 °C by Soxhlet extractor. The solvent was evaporated by a rotary evaporator. The obtained oil was esterified to determine the fatty acid composition. The extraction yield was found as 1.13 %.

Preparation of fatty acid methyl esters (FAMEs): The fatty acids were esterified into methyl esters by saponification with 0.5 N methanolic sodium hydroxide and transesterified with 14 % (v/v) boron trifluoride (BF₃) in methanol under reflux³³.

GC conditions: Fatty acid methyl esters (FAMEs) were analyzed on a HP (Hewlett Packard) Agilent 6890 N model gas chromatograph (GC), equipped with a flame ionization detector (FID) and fitted to a Supelco SP-2380 Fased Silica

capillary column (60 m, 0.25 mm i.d. and 0.2 μ m). Injector and detector temperatures were set at 250 and 260 $^{\circ}$ C, respectively. The oven was programmed at an initial temperature of 140 $^{\circ}$ C and an initial time of 5 min. Thereafter the temperature was increased up to 240 $^{\circ}$ C at a rate of 3 $^{\circ}$ C min $^{-1}$. The total run time was 41.33 min. Helium was used as the carrier gas (1 mL min $^{-1}$). Identification of fatty acids was carried out by comparing sample FAME peak relative retention times. The results were expressed as FID response area in the relative percentages. Each reported result is given as the average value of three GC analyses. The results are offered as means \pm S.D.

Essential oil analysis: Hydrodistillation of the air-dried plant material for 3 h using a Clevenger-type apparatus afforded a yellow essential oil. The obtained essential oil was stored at + 4 $^{\circ}$ C until use. The total yield was 0.05 % (w/w).

GC and GC/MS conditions: The GC/MS analysis were carried out with an Agilent 5975 GC-MSD system. Innowax FSC column (60 m \times 0.25 mm, 0.25 mm film thickness) was used with helium as carrier gas (0.8 mL/min). GC oven temperature was kept at 60 $^{\circ}$ C for 10 min and programmed to 220 $^{\circ}$ C at a rate of 4 $^{\circ}$ C/min and kept constant at 220 $^{\circ}$ C for 10 min and then programmed to 240 $^{\circ}$ C at a rate of 1 $^{\circ}$ C/min. Split ratio was adjusted at 40:1. The injector temperature was set at 250 $^{\circ}$ C. Mass spectra were recorded at 70 eV. Mass range was from m/z 35 to 450.

The GC analysis was carried out using an Agilent 6890N GC system. FID detector temperature was 300 $^{\circ}$ C. To obtain the same elution order with GC-MS, simultaneous auto-injection was done on a duplicate of the same column applying the same operational conditions. Relative percentage amounts of the separated compounds were calculated from FID chromatograms. The analysis results are given in Table-1.

Identification of components: Identification of the essential oil components were carried out by comparison of their relative retention times with those of authentic samples or by comparison of their relative retention index (RRI) to series of *n*-alkanes. Computer matching against commercial (Wiley GC/MS Library, Adams Library, MassFinder 3 Library)^{34,35} and in-house "Baser Library of Essential Oil Constituents" built up by genuine compounds and components of known oils, as well as MS literature data^{36,37} was used for the identification.

Biological activity assays

Preparation of plant extracts: Chloroform and ethanol extracts were separately prepared from 40 g batches of the air-dried and powdered plant materials by extracting with 400 mL solvent at room temperature under stirring for 24 h. Then the solvents were evaporated to dryness in vacuo (60 $^{\circ}$ C). The yields of chloroform and ethanol extracts of *R. pedatus* subsp. *pedatus* were 1.06 and 2.12 %, respectively. All the extracts were stored at -20 $^{\circ}$ C.

Cytotoxic activity assay

Cell lines and culture medium: PC3, Vero and HeLa cell lines were obtained from ATCC. All cells were cultivated in a humidified incubator at 37 $^{\circ}$ C with 5 % CO $_2$. Cells were cultured in DMEM supplemented with L-glutamine (2 mmol/L) and 10 % fetal bovine serum. All the tissue culture reagents were purchased from Biological Industries (Israel).

Determination of cell viability by Real-Time Cell Analyzer

(RTCA): Cells were grown and expanded in 100 mm tissue culture dishes. After reaching 60-80 % confluence, cells were washed with PBS and detached from the flasks by trypsin/EDTA treatment. Meanwhile, 100 μ L of cell culture media at room temperature was added into each well of E-plate 96 and background of E-plate was measured. To determine the optimum cell number, 5000 cells/well were seeded for each cell line. After 0.5 h of incubation at room temperature, E-plates were placed into the Real-Time Cell Analyzer MP (RTCA) station. Cells were grown and impedance was measured every 0.5 h as the cell index (CI) value. To determine the effect of test extracts, optimum number of cells (PC3:7500, HeLa: 7500 and Vero: 7500) for each cell line were seeded. After 16-24 h cells were exposed to test extracts at different concentrations (25, 100, 250, 500 and 1000 μ g/mL). CI values were monitored every 2 min for 2 h to visualize the fast drug response and then every 0.5 h for the late drug response. The electrical impedance was measured by RTCA software of the xCELLingence system as a dimensionless parameter termed CI. All the measurements were done at least in triplets and IC $_{50}$ values were determined using RTCA software.

DPPH radical scavenging assay: DPPH (1,1-diphenyl-2-picrylhydrazyl; Fluka, Steinheim, Germany) radical scavenging activity of *R. pedatus* subsp. *pedatus* was evaluated according to a modified version of the method described by Gezer *et al.*³⁸. One thousand μ L of 1 mg/mL concentration of the extracts in ethanol and chloroform were added to 4 mL of 0.004 % methanol solution of DPPH. After a 0.5 h incubation period at room temperature, the absorbance was read against a blank at 517 nm. Inhibition of free radical by DPPH was calculated by the following equation:

$$I \% = [(A_b - A_s) / A_b] \times 100$$

where A_b is the absorbance of the control reaction and A_s is the absorbance of the test sample. α -tocopherol (TOC) was used for comparison. Tests were carried out in triplicate.

Determination of total phenolic and flavonoid contents:

Total phenolic constituents was determined by using the Folin-Ciocalteu method³⁹. 0.1 mL of the extracts (0.5 and 1 mg/mL) were mixed with 2.8 mL deionized water. This solution was mixed with 2 mL 2 % sodium carbonate and 0.1 mL of 0.1 N Folin-Ciocalteu reagent. After incubation at room temperature for 0.5 h the absorbance of the mixture was measured at 750 nm against a deionized water blank on a UNIQCAM 8625 UV/visible spectrophotometer. Gallic acid was chosen as a standart. The data expressed as miligram gallic acid equivalents.

Total flavonoid content was determined by the aluminium chloride colorimetric method described by Chang *et al.*⁴⁰ 0.5 mL of the extracts (0.5 and 1 mg/mL) were mixed with 1.5 mL of ethanol, 0.1 mL of 10 % aluminium chloride and 2.8 mL of distilled water. The mixture was kept at room temperature for 0.5 h and the absorbance was recorded at 415 nm with the UNIQCAM 8625 UV/visible spectrophotometer. Quercetin equivalent was chosen as a standard. The amount of flavonoid was expressed as equivalent.

RESULTS AND DISCUSSION

The results of essential oil analysis of *R. pedatus* subsp. *pedatus* is shown in Table-1. GC/MS analysis revealed 29

components representing 99.1 % of essential oil. Protoanemonin (14.3 %), phytol (13.7 %), (Z)-3-hexenal (7.8 %), octanal (6.7 %) and a sesquiterpenoid compound α -bisabolol (7.9 %) were found to be the main components in the essential oil from the aerial parts of *R. pedatus* subsp. *pedatus*. Phytol, showing similarity to our results, was also previously found to be the main components of the essential oil of *R. constantinopolitanus* (23.6 %) and *R. arvensis* (19.5 %)³².

TABLE-1
CHEMICAL CONSTITUENTS OF ESSENTIAL OIL FROM *R. pedatus* subsp. *pedatus*

RRI	Compounds	%
1048	2-Methyl-3-buten-2-ol	1.2
1093	Hexanal	2.9
1225	(Z)-3-Hexenal	7.8
1296	Octanal	6.7
1304	1-Octen-3-one	1.6
1327	(Z)-3-Hexenyl acetate	1.3
1391	(Z)-3-Hexenol	1.5
1400	Nonanal	2.2
1452	1-Octen-3-ol	0.7
1532	Camphor	1.5
1541	Benzaldehyde	0.7
1553	Linalool	1.2
1596	Protoanemonin*	14.3
1600	Hexadecane	0.5
1663	Phenylacetaldehyde	2.1
1726	Germacrene D	0.7
1758	(E,E)- α -Farnesene	0.7
1765	Geranyl acetate	2.3
1857	Geraniol	6.0
1958	(E)- β -Ionone	2.3
2041	Pentadecanal	3.3
2131	Hexahydrofarnesylacetone	0.5
2200	Docosane	0.8
2242	Methyl hexadecanate	0.3
2232	α -Bisabolol	7.9
2300	Tricosane	5.8
2500	Pentacosone	5.0
2622	Phytol	13.7
2931	Hexadecanoic acid	3.6
Total		99.1

RRI: Relative retention indices calculated against n-alkanes, % calculated from FID data, tr Trace (< 0.1 %), *Tentative identification

The fatty acid composition of *R. pedatus* subsp. *pedatus* is given in Table-2. Twenty seven fatty acids were identified by using GC and GC-MS. Linoleic acid (24.47 %) and palmitoleic acid (18.94 %) were found as main constituents of *Ranunculus pedatus* subsp. *pedatus*. Unsaturated fatty acids amounted to 60.43 % of the total fatty acids, while the saturated fatty acids were 17.83 %. The presence of linoleic acid in adequate amounts is essential, because it is an essential fatty acid. Lack of dietary essential fatty acids such as linoleic acid has been implicated in aetiology of diseases including cardiovascular disease and its progression⁴¹. Linoleic acid can not be synthesized by the human body and is very important for the nutritional value of oils⁴².

The results of the cytotoxic activity are shown in Table-3. Ethanol and chloroform extracts of *R. pedatus* showed strong effect on HeLa with IC₅₀ values of 47.97 and 78.48 μ g/mL, respectively. In a previous report, Lorimer *et al.*⁴³ evaluated

cytotoxic activity of ethanol extract of *Ranunculus pinguis* against P388 cell line. The extract of *R. pinguis* was shown to have strong inhibitory activity. They reported that the cytotoxic activity of *R. pinguis* against P388 cell line was due to the presence of a glucoside, ranunculin.

TABLE-2
FATTY ACID COMPOSITION OF *R. pedatus* subsp. *pedatus* (%)

Fatty acids	Content (%)
C6:0	0.10 \pm 0.01 ^a
C8:0	0.06 \pm 0.02
C10:0	0.09 \pm 0.01
C11:0	0.09 \pm 0.02
C13:0	0.11 \pm 0.01
C15:0	0.26 \pm 0.03
C16:0	2.78 \pm 0.02
C17:0	0.46 \pm 0.02
C18:0	0.39 \pm 0.01
C20:0	12.71 \pm 0.01
C21:0	0.19 \pm 0.03
C23:0	0.59 \pm 0.03
ΣSFA^b	17.83 \pm 0.22
C14:1 ω 5	0.80 \pm 0.02
C16:1 ω 7	18.94 \pm 0.01
C17:1	0.41 \pm 0.02
C18:1t	3.96 \pm 0.01
C 18:1 ω 9	1.01 \pm 0.01
C20:1	0.27 \pm 0.03
C24:1 ω 9	0.36 \pm 0.01
ΣMUFA^b	25.75 \pm 0.11
C18:2n6t	5.41 \pm 0.05
C18:2 ω 6	24.47 \pm 0.02
C20:2 ω 6	0.13 \pm 0.01
C22:2	0.06 \pm 0.02
C18:3 ω 6	1.25 \pm 0.01
C20:3 ω 6	1.43 \pm 0.01
C20:4 ω 6	0.17 \pm 0.02
C20:5 ω 5	1.76 \pm 0.01
ΣPUFA^b	34.68 \pm 0.15

^aAverage of three lots analysed, ^bValues reported are means \pm SD. SFA: Saturated fatty acids, MUFA: Monounsaturated fatty acids, PUFA: Polyunsaturated fatty acids

TABLE-3
CYTOTOXIC ACTIVITY RESULTS OF EXTRACTS OF *R. pedatus* subsp. *pedatus*

Extract	IC ₅₀ (μ g/mL) ^a	IC ₅₀ (μ g/mL) ^a	IC ₅₀ (μ g/mL) ^a
	Pc3	HeLa	Vero
Chloroform	78.48	NA ^b	NA
Ethanol	47.97	174.04	144.62

^aValues are mean triplicate analysis, ^bNA: Not active

The presence of flavonoids (quercetin, kaempferol, vitexin, isovitexin, orientin and isoorientin)¹¹⁻¹⁵, alkaloids (berberine, palmatine, columbamine and magnoflorine)^{18,19} and triterpenes (ursolic acid, oleanolic acid and betulinic acid)¹⁶ were previously reported by phytochemical studies on various species of *Ranunculus*. Vitexin, isovitexin, *p*-hydroxy benzoic acid and *p*-coumaric acid from *R. pedatus* subsp. *pedatus* were isolated and reported in our previous work. The cytotoxic activities of the fractions were also evaluated by the brine shrimp lethality bioassay. The moderate cytotoxic activity was attributed to presence of vitexin and isovitexin¹⁵ which were previously found to be cytotoxic⁴⁴. Cytotoxic effect of berberine was also

determined against human tumor cell lines⁴⁵. Ursolic acid was shown to have moderate cytotoxicity against human cancer lines⁴⁶. Vitexin, isovitexin, ursolic acid and berberine might be responsible for the observed cytotoxic activities of investigated extracts.

The antioxidant activity of the ethanol and chloroform extracts of *R. pedatus* were reported in Table-4. The ethanol extract showed 67.54 % DPPH inhibition. A positive correlation between flavonoid and total phenolic contents of the extracts and the antioxidant activity was observed. The TPC of the ethanol and chloroform extracts of *R. pedatus* subsp. *pedatus* were determined as 118.4 and 472.06, gallic acid equivalents (mg/g) dry matter. TFC of the ethanol extract of *R. pedatus* subsp. *pedatus* was obtained as 4.36 %, whereas chloroform extract had only 1.23 %. Typical phenolics that possess antioxidant activity have been characterized as flavonoids and phenolic acids⁴⁷. Phenols and polyphenols known as chain-breaking antioxidants are very important constituents of plants⁴⁸. Many flavonoids were isolated in some *Ranunculus* species in previous studies¹²⁻¹⁵. Vitexin and isovitexin were reported to have antioxidant activity in a previous study⁴⁹. Vitexin and isovitexin could be responsible for antioxidant activity observed for the ethanol extract of *R. pedatus* subsp. *pedatus*. Kaya *et al.* studied the flavonoid and phenolic content and antioxidant activities of hexane, ethyl acetate, methanol and water extracts of *R. marginatus* var. *trachycarpus* and *R. sprunerianus* and highest antioxidant activity was observed for methanol extracts of the plant²⁴. In addition, the antioxidant activity of aqueous methanolic extract of *R. repens* flowers and leaves were 0.12 mM TE/g and 0.13 mM TE/g, respectively²⁷.

TABLE-4
DETERMINATION OF ANTIOXIDANT ACTIVITY,
TOTAL PHENOLIC AND FLAVONOID CONTENT
OF *R. pedatus* subsp. *pedatus*

Extract	DPPH (%)	TPC (mg/g) ^a	TFC (g %) ^b
Chloroform	31.82 ± 4.04 ^c	118.4 ± 5.74	1.233 ± 0.05
Ethanol	67.54 ± 5.49	472.06 ± 3.72	4.368 ± 0.12

^aTPC: Total phenolic content (mg/g dry mass), ^bTFC: Total flavo-noid content (g %), ^cResults are mean ± SD of three replicate analysis

Conclusion

In conclusion, this is the first report on the fatty acid and essential oil composition and the cytotoxic and antioxidant activities of *R. pedatus* subsp. *pedatus* from Turkey²⁹. Components were identified in the essential oil and the main components were phytol and protoanemonin. The main constituents of fatty acid of *R. pedatus* subsp. *pedatus* were linoleic and palmitoleic acid. It is clear that, there is a significant correlation between the total phenolic and flavonoid content and antioxidant activity. Further studies are required for the isolation and structure elucidation of individual compounds responsible for the observed biological activities.

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